

Synthesis of a Biologically Active Vitamin-D₂ Metabolite

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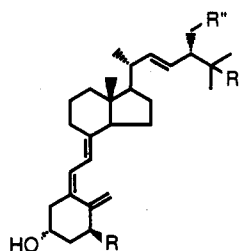
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The synthesis of 1 α ,25,28-trihydroxyvitamin-D₂ (**1**) from vitamin-D₂ is described. Approaches to the upper side-chain synthon via the chiral sulfone **15**, prepared either through the enzymatic resolution of **10** or the opening of the optically pure epoxide **17** with the dianion of methyl phenyl sulfone, are described. A Julia coupling of the sulfone **15** with C-1 hydroxylated aldehyde **7**, obtained from vitamin-D₂, gave the (2*E*)-isomer **24a**, which was deprotected and photoisomerized to give **1**.

Introduction

1 α ,25,28-Trihydroxyvitamin-D₂ [1,25,28-(OH)₃-D₂] **1** has been tentatively identified¹ as an active metabolite of vitamin-D₂. Structure **1** was proven unequivocally by a total synthesis by Batcho² in which the two halves of the molecule were prepared in optically pure form and joined at C7-C8 by a Wittig-Horner reaction. This metabolite



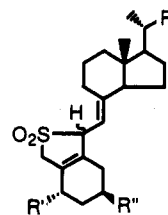
- 1** R,R',R'' = OH 1,25,28 (OH)₃ Vitamin-D₂
2 R, R', R'' = H Vitamin-D₂

has antiproliferative activity against keratinocytes and sebocytes and has low calcium liability.³ At 10⁻⁶ M in vitro, **1** changed the relative number of basal, squamous, and envelope-type keratinocytes² from 98:17:15 (control) to 51:26:29. We report herein a novel synthesis of **1** using vitamin-D₂ itself as the starting material. The synthesis (Scheme I) involves three key features: (i) the dissection of the vitamin-D₂ side chain, (ii) C-1 hydroxylation, and (iii) the preparation and introduction of the appropriate upper side chain. The latter was accomplished via a Julia coupling of the aldehyde **7** with the chiral sulfone **15**.

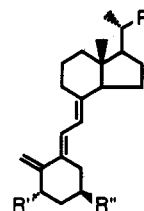
Results and Discussion

Our approach was based on the assumption that aldehyde **7** was readily accessible in large quantities and would be an ideal late-stage intermediate. Aldehyde **7** was prepared from vitamin-D₂ (ergocalciferol, **2**) with some modification of the procedures described by Calverley.⁴ Treatment of **2** in CH₂Cl₂ with SO₂ at -10 °C, followed by

silylation with *tert*-butyldimethylsilyl chloride, gave **3b** as a free-flowing solid. Cheletropic extrusion of SO₂ from **3b** followed by hydroxylation at C-1 with selenium dioxide in the presence of 4-methylmorpholine *N*-oxide (NMO), as described by Barton⁵ et al., gave a 7:1 ratio of 1 α /1 β epimeric alcohols, which after filtration to remove polymeric material, was silylated and crystallized to give the (5*E*)-1 α -hydroxy derivative, **4c**. Attempts to effect C-1 hydroxylation of the natural (5*Z*) isomer TBDMS (protected **2**) with SeO₂ were unsuccessful. The triene unit in **4c** was again protected with SO₂, and the resulting sulfone was ozonized. Extrusion of SO₂ gave the aldehyde **7**. NMR analysis of crude **7** indicated ca. 10% of the C-20(R) isomer to be present but it could be removed by chromatography.



- 3a** R = ; R' = H; R'' = OH
3b R = ; R' = H; R'' = OTBDMS
5 R = ; R', R'' = OTBDMS
6 R = CHO; R', R'' = OTBDMS



- 4a** R = ; R' = H, R'' = OTBDMS
4b R = ; R' = OH, R'' = OTBDMS
4c R = ; R', R'' = OTBDMS
7 R = CHO; R', R'' = OTBDMS

The first attempt to prepare an optically active C₆ synthon for the side chain was by enzymatic resolution of

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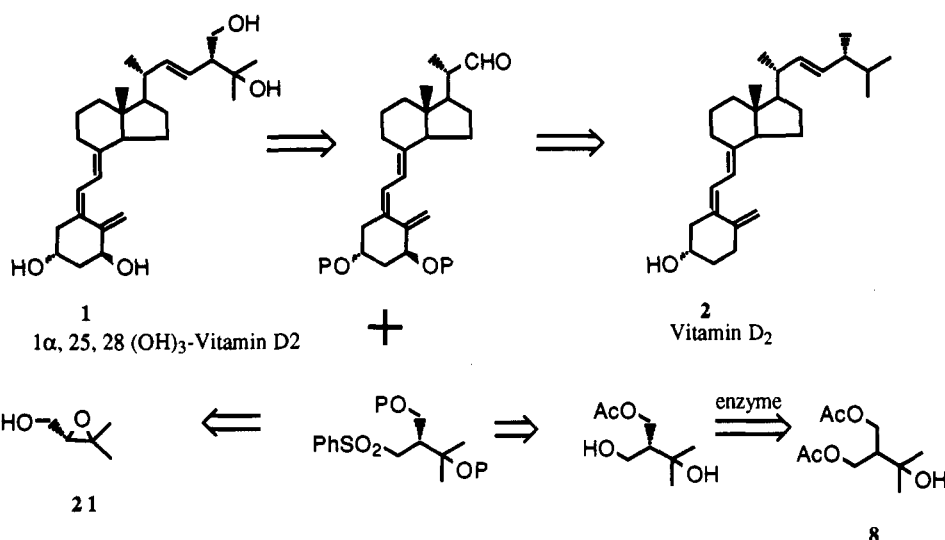
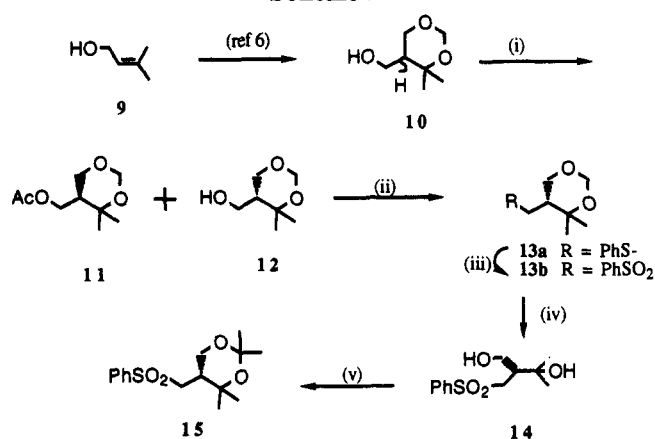
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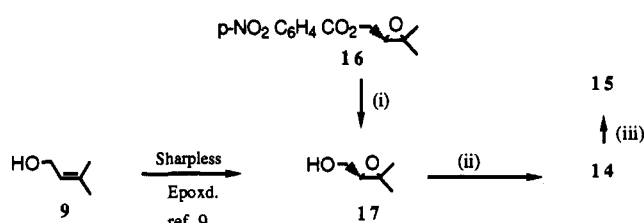
Scheme I

Scheme II^a

^a Key: (i) porcine pancreatic lipase; (ii) diphenyl disulfide/tri-*n*-butylphosphine; (iii) *m*-CPBA/CH₂Cl₂; (iv) boron trichloride/CH₂Cl₂; (v) 2,2-dimethoxypropane/PhH/cat. TsOH.

a suitable symmetrical compound such as 8 (Scheme I) or its derivative. The preparation of the backbone of 8 is outlined in Scheme II. A Prins reaction between 3-methyl-2-buten-1-ol and formaldehyde provided the known alcohol⁶ 10. Lipase from *P. fluorescens* showed no selectivity in acetylation toward 10, but lipase from *Candida cylindracea* and especially porcine pancreatic lipase showed significant selectivity. The L-isomer was acetylated preferentially with concomitant enrichment of the desired *R*-isomer. From 70 g of 10, we obtained 21 g of unreacted *R*-isomer (12) with an enantiomeric purity of better than 95%. The two-step conversion of the hydroxyl group of 12 to the phenylsulfonyl group proceeded uneventfully and provided 13b. The absolute configuration of 13b as depicted was established by X-ray crystallographic analysis.

The methylidene acetal in 13b proved to be extremely acid stable so its use as a protective group slated for eventual removal in the presence of the sensitive triene unit of the vitamin-D system could not be contemplated. Hence, an easily removable protecting group such as disilyl or the acetonide group was sought. Attempts at direct transacetalization of 13b to 15 with 2,2-dimethoxypropane

Scheme III^a

^a Key: (i) aqueous NaOH/MeOH; (ii) CHSO₂Ph²⁻ (18)/THF; (iii) 2,2-dimethoxypropane/PhH/cat. TsOH.

using a variety of acidic conditions were unsuccessful. However, the methylidene group was easily removed when 13b was exposed to BCl₃ (1.0 equiv in CH₂Cl₂ at -78 °C) to furnish diol 14 in 30% yield. It appears that Me₂BCl is more promising and gives a better yield (44%) in this conversion. Conversion of the diol to the acetonide worked well using either 2,2-dimethoxypropane or 2-methoxypropane and gave the acetonide sulfone 15 in 70% yield. The reaction using the conventional acetone/copper sulfate method did not occur presumably because of the hindered nature of the diol.

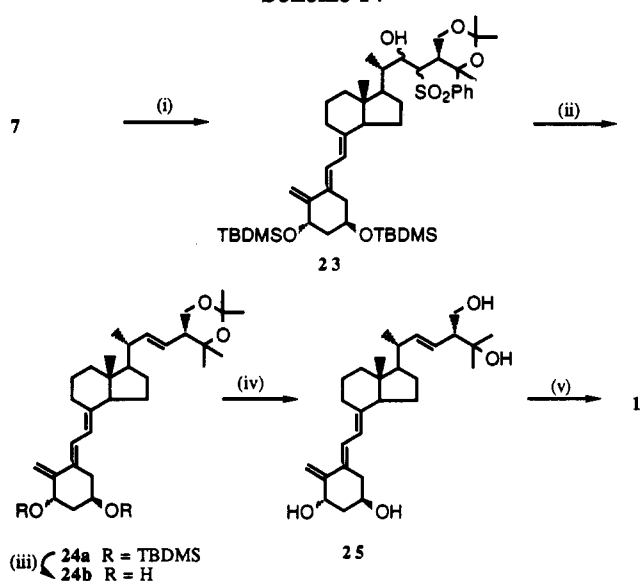
Another approach for the preparation of chiral sulfone 15, which ultimately proved to be most expeditious, was through the ring opening⁷ of epoxide 17 with the dianion of methyl phenyl sulfone,⁸ 18 (Scheme III). The known chiral epoxide has been prepared⁹ through the Sharpless epoxidation of isoprenyl alcohol 9. The epoxide is also available commercially as its *p*-nitrobenzoate ester (16) which we chose as starting material. Ester 16 was conveniently hydrolyzed by addition of 1 equiv of aqueous sodium hydroxide in methanol. To prevent rearrangement of the epoxy alcohol, the free hydroxyl group was protected as its silyl derivative 19 before being allowed to react with 18. The reaction, however, was very sluggish and under harsh conditions afforded only the undesired product

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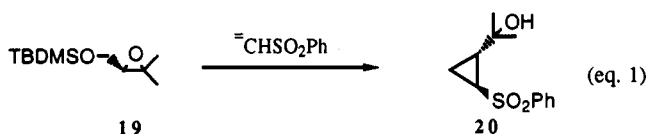
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Scheme IV^a

^a Key: (i) 15 + 1 equiv of LDA and then 7; (ii) Na-Hg/MeOH/phosphate buffer; (iii) (*n*-Bu)₄N⁺F⁻/THF; (iv) Dowex 50W-X4 ion-exchange resin/methanol; (v) 450-W medium-pressure lamp/MeOH.

cyclopropyl derivative 20 as a single isomer which was tentatively assigned the *trans* configuration as depicted (eq 1). When unprotected 17 was exposed to 18, the



reaction occurred smoothly at room temperature to give the desired sulfone diol 14 with the free hydroxyl group apparently assisting the opening of the epoxide (presumably through lithium coordination¹⁰). No product resulting from the rearrangement of epoxide 17 prior to its opening was observed.

A Julia coupling¹¹ of 7 with the anion derived from sulfone 15 (Scheme IV) gave a diastereomeric mixture of hydroxy sulfones 23, reduction of which with sodium amalgam¹² in the presence of sodium hydrogen phosphate buffer, gave the 22*E* product 24a in 25% yield. The desilylation of 24a with tetrabutylammonium fluoride followed by removal of the acetonide group gave 25, which on photoisomerization gave 1.

Experimental Section

General Methods. Melting points were determined in capillaries and are uncorrected. Unless otherwise indicated, IR and NMR were determined in CHCl₃ and CDCl₃, respectively; ¹H NMR spectra were recorded at 200 and/or 400 MHz. Mass spectra (MS) were determined with a direct inlet system with ionization energy of 70 eV; *m/z* values are given with relative intensities in parentheses. TLC plates (silica gel G) were purchased from E. Merck (Darmstadt); spots were visible under short-wavelength UV light and were made visible to the eye by spraying with 10% phosphomolybdic acid in ethanol and heating to 100 °C. Chromatographic purifications were carried out with

Fluka silica gel 60 (220–440 mesh unless otherwise stated). All chromatographed products were homogeneous by silica gel TLC.

SO₂-Adduct of 3(*S*)-[*tert*-Butyldimethylsilyloxy]-9,10-secoergosta-5,7(*E*),10(19),22(*E*)-tetraene (3b). A 1-L, three-necked, round-bottomed flask equipped with a mechanical stirrer, Ar inlet, and dry ice condenser was cooled to -25 °C, and SO₂ (130 g) was condensed into it. A solution of 100 g (0.252 mol) of vitamin D₂ (2) in CH₂Cl₂ (250 mL) was added. The resulting mixture was stirred at -10 °C for 1.25 h then carefully evaporated at 30 °C. The CH₂Cl₂ was removed at 45 °C using a water aspirator to give a dark brown gel. This was dissolved in CH₂Cl₂ (350 mL) cooled to 5 °C and, with stirring under argon, treated with imidazole (22.5 g, 0.33 mol) and TBDMS chloride (50 g, 0.33 mol). The resulting heterogeneous mixture was stirred at rt overnight and then filtered through Celite. The Celite was washed with CH₂Cl₂ (2 × 250 mL), and the combined washings and filtrate were washed with brine (500 mL), dried (MgSO₄), and evaporated to give 140.2 g (97%) of 3b as a pale yellow solid. Isolation and characterization of the 6*S* and 6*R* isomers have been reported.⁴

TBDMS Ether of 5(*E*)-Vitamin-D₂ (4a). A stirred mixture of 3b (140.0 g, 0.243 mol) and NaHCO₃ (140 g) in 95% EtOH (1.0 L) was boiled under reflux for 1.25 h, and 750 mL of EtOH was removed by distillation (water aspirator). Hexane (1 L) was added and the resulting mixture was stirred at reflux for 15 min, cooled to rt, and filtered. The filter cake was washed with hexane (2 × 200 mL), and the combined filtrate and washings were washed with brine (1.5 L), dried (MgSO₄), and evaporated to give 117.2 (94% yield) of 4a as a pale yellow gum. TLC (25% CH₂Cl₂ in hexane) showed the product at *R*_f = 0.43. An analytical sample was obtained by flash chromatography over silica gel with 1% EtOAc in hexane as eluent gave 4a as a gum: [α]_D²⁵ +87.7° (CHCl₃, *c* = 0.9508); UV (EtOH) λ_{max} 271 (ϵ = 20 600) nm; ¹H NMR δ 0.065 (6 H, s, Me₂Si), 0.57 (3 H, s, CH₃-18), 0.85 (6 H, d, *J* = 7 Hz, CH₃-26 + CH₃-27), 0.89 (9 H, s, *t*-BuSi), 0.91 (3 H, d, *J* = 6 Hz, CH₃-28), 1.02 (3 H, d, *J* = 6 Hz, CH₃-21), 3.84 (1 H, m, CH-3), 4.64 (1 H, s, CH_A-19), 4.93 (CH_B-19), 5.20 (2 H, dd, *J* = 5, 15 Hz, CH-22 + CH-23), 5.90 (1 H, d, *J* = 14 Hz, CH-7), 6.46 (1 H, d, *J* = 14 Hz, CH-6). Anal. Calcd for C₃₄H₅₈OSi: C, 79.93; H, 11.44. Found: C, 80.28; H, 12.15.

1(*S*),3(*R*)-Bis(*tert*-butyldimethylsilyloxy)-9,10-secoergosta-5(*E*),7(*E*),10(19),22(*E*)-tetraene (4c). A mixture of pulverized SeO₂ (6.296 g, 0.0567 mol) and CH₃OH (600 mL) was boiled under reflux until a solution was obtained (ca. 20 min), and then the solution was cooled to 30 °C and treated with a solution of 4a (116.0 g, 0.227 mol) followed immediately by NMMO (53.19 g, 0.454 mol). The resulting mixture was stirred at reflux for 5.5 h, cooled to rt, and poured into a mixture of CH₂Cl₂ (1 L) and brine (1.5 L). The organic phase was separated, and the aqueous phase was extracted with CH₂Cl₂ (1.5 L). The combined organic extract was washed with 5% NaHCO₃ (1.5 L) and 10% brine (2.4 L), dried (MgSO₄), and evaporated to give 134 g of a gum. This was chromatographed over 1.5 kg of silica gel with 8.0 L of 2.5% EtOAc in hexane (to remove nonpolar material) followed by 16.0 L of 75% EtOAc in hexane. Evaporation of the appropriate fractions gave 54.9 g of a gum. This was dissolved in CH₂Cl₂ (220 mL), cooled to 10 °C, and treated with imidazole (9.3 g, 0.137 mol) and TBDMSCl (20.6 g, 0.137 mol). The resulting mixture was stirred at rt for 6 h and filtered through Celite, and the filter cake was washed with CH₂Cl₂ (100 mL). The combined filtrate and washings were washed with brine (3 × 100 mL), dried (MgSO₄), and evaporated to a semisolid. This was dissolved, with stirring, in ethyl acetate (175 mL), warmed to 70 °C, and diluted with MeOH (220 mL). After being cooled to room temperature, the product (41.56 g) crystallized and was collected by filtration. Recrystallization from EtOAc-MeOH gave 40.42 g (28% overall) of 4c: mp 115–116 °C (lit.⁴ mp 113–114 °C); [α]_D²⁵ +62.45 (CHCl₃, *c* = 0.9802). The UV, NMR, and MS data agree with those reported.⁴

(6*S*)- and (6*R*)-SO₂ Adducts of 4c (5). A solution of 4c (50 g) in CH₂Cl₂ (100 mL) was added to SO₂ (100 mL) at -10 °C with stirring. The mixture was stirred for 45 min and evaporated to give 63.8 g of the known⁴ sulfone adducts (78:22 mixture of diastereomers) as a foam.

(6*S*)- and (6*R*)-SO₂ Adducts of 1(*S*),3(*R*)-Bis(*tert*-butyldimethylsilyloxy)-20(*S*)-formyl-9,10-secopregna-5,7(*E*),10(19)-triene (6). A portion of the above-prepared SO₂ adducts

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(51.1 g, 72.5 mmol) was dissolved in CH₂Cl₂ (380 mL) and MeOH (130 mL). The resulting solution was cooled to -10 °C and, with stirring, treated with O₃ (O₂ pressure = 8 psi) for 40 min, followed by Ar for 10 min. Ph₃P (25 g, 95.4 mmol) was added portionwise at a rate such that the temperature was kept below 0 °C. The mixture was kept at rt for 3 h and then evaporated. The residue was dissolved in ether (150 mL), hexane (150 mL) was added, and the precipitated Ph₃PO was collected by filtration. Evaporation of the filtrate and chromatography of the residue over 480 g of silica gel with an increasing concentration of EtOAc in hexane (1–15%) and collection of the desired fractions gave 33.3 g (78% yield from 5) of 6 as a colorless solid. The major isomer (6S) was obtained as needles: mp 117–118 °C (lit.⁴ mp 122–124 °C); [α]_D²⁵ +9.21° (CHCl₃, c = 1.041). The minor (6R, more polar) isomer was obtained as a gum, [α]_D²⁵ -18.54° (CHCl₃, c = 0.8898).

1(S),3(R)-Bis[(*tert*-butyldimethylsilyloxy)-20(S)-formyl-9,10-secopregna-5(E),7(E),10(19)-triene (7). A solution of 6 (28.85 g, 45.3 mmol) in 95% ethanol (250 mL) was treated with NaHCO₃ (28.4 g, 0.34 mol), and the mixture was stirred at reflux for 1.5 h. It was cooled to rt, diluted with EtOAc (100 mL), and filtered through Celite, which was washed with EtOAc. Evaporation of the combined filtrates followed by chromatography of the residue over 258 g of silica gel gave 16.39 g (63%) of 7: mp 114–116 °C (lit.⁴ mp 113–115 °C); [α]_D²⁵ +55.20° (CHCl₃, c = 1.098). The IR, UV, NMR, and MS data agree with those reported.⁴

(R)-4,4-Dimethyl-1,3-dioxane-5-methanol (12). Racemic alcohol 10 (71.25 g, 0.487 mol, 95% pure by GC) was added under Ar to a magnetically stirred suspension of vinyl acetate (850 mL) and lipase (100 g, Sigma Lot 39FO454, type II crude porcine pancreatic lipase) in a 2-L round-bottomed flask. The progress of the transesterification was monitored by GC on a fused-silica capillary column with permethylated β-cyclodextrin as stationary phase (0.32 mm × 25 m) at a temperature gradient of 4 °C/min, 100–200 °C within 50 min, and a H₂ flow rate of 52 cm³/s. Under these conditions, racemic 10 was resolved with retention times of 34.95 min for desired isomer 11 and 38.33 min for the dextrorotatory enantiomer. After 8.5 h, the reaction was 75% complete and GC showed 12 as a single enantiomer. The separation of 12 from the acetate 11 was achieved by HPLC on a stainless steel column (10 cm × 96 cm) packed with silica gel (40 nm) at a flow rate of 250 mL/min with RI detection and using 1:4 ethyl acetate–hexane followed by 1:1 EtOAc–hexane as eluent. This provided 56.87 g of acetate 11 and 18.58 g (26%) of enantiomerically enriched alcohol 12 as a colorless liquid: [α]_D²⁵ -15.95° (c = 1.17%, CHCl₃); ¹H NMR δ 1.23, 1.36 (3 H, s, s, (CH₃)₂C), 1.44 (1 H, br t, OH), 1.95 (1 H, m, CH), 3.52, 3.74 (2 m, 1 H each, CH₂OH), 3.72, 4.09 (2 dd, 1 H each, *J*_{vic} = 8.9 and 4.3 Hz, *J*_{gem} = 11.6 Hz, CH₂O), 4.87 (2 H, AB, OCH₂O); MS 145 (<1, M-1), 131 (4), 101 (5), 81 (3), 71 (17), 69 (11), 59 (100). Anal. Calcd for C₉H₁₆O₄: C, 57.51; H, 9.65. Found: C, 57.21; H, 9.96.

4,4-Dimethyl-5-[(phenylthio)methyl]-1,3-dioxane (13a). A mixture of (2.92 g, 20.0 mmol) of 12 (2.92 g, 20.0 mmol) and PhSSPh (5.2 g, 24 mmol) was taken up in dry DMF (20 mL) to which was then added dropwise *n*-Bu₃P (4.85 g, 24 mmol). The resulting mixture was stirred at room temperature overnight and then subjected to extractive workup with ether, drying (Na₂SO₄), and evaporation. The crude product was chromatographed on silica gel to give 3.6 g (76% yield) of 13a, as a thick colorless oil: [α]_D²⁵ -85.62° (CHCl₃, c = 0.537); ¹H NMR δ 1.25, 1.29 (2 s, 3 H each, (CH₃)₂C), 2.00 (1 H, m, CH), 2.51, 3.06 (dd, *J*_{vic} = 11.4 and 3.1 Hz, *J*_{gem} = 13.2 Hz, CH₂S), 3.59, 4.19 (2 dd, 1 H each, *J*_{vic} = 9.9 and 4.3 Hz, *J*_{gem} = 11.7 Hz, CH₂O), 4.84 (2 H, AB, OCH₂O), 7.21, 7.30, 7.35 (1 H, 2 H, 2 H, C₆H₅). Anal. Calcd for C₁₃H₁₈SO₂: C, 65.51; H, 7.61. Found: C, 65.79; H, 7.64.

4,4-Dimethyl-5-[(phenylsulfonyl)methyl]-1,3-dioxane (13b). To a stirred solution of 13a (27.7 g, 116 mmol) in CH₂Cl₂ (200 mL) at 0 °C was added portionwise a suspension of 85% *m*-CPBA (55 g) in CH₂Cl₂ (600 mL). The mixture was stirred for 0.5 h at 0 °C and then for 24 h at rt, diluted with CH₂Cl₂ (100 mL), and filtered. The filtrate was washed with aqueous 7% NH₃ and then dried (Na₂SO₄), filtered, and evaporated to give 33.04 g of 13b as a white solid. Chromatography over silica gel column eluting with 3:2:5 EtOAc–CH₂Cl₂–hexane gave 30.55 g (97%) of 13b as crystalline solid: mp 144–145 °C; [α]_D²⁵ -28.30°

(CHCl₃, c = 0.554); ¹H NMR δ 1.15, 1.16 (2 s, 3 H each, (CH₃)₂C), 2.94, 3.02 (2 dd, 1 H each, *J*_{vic} = 9.8 and 1.8 Hz, *J*_{gem} = 14.6 Hz, CH₂SO₂), 3.70, 4.20 (2 dd, 1 H each, *J*_{vic} = 8.9 Hz and 4.2 Hz, *J*_{gem} = 11.9 Hz, CH₂O), 4.83 (2 H, s, OCH₂O), 7.61 (2 H, t, *m*-ArH), 7.70 (1 H, t, *p*-ArH), 7.94 (2 H, d, *o*-ArH). Anal. Calcd for C₁₃H₁₈SO₄: C, 57.76; H, 6.71. Found: C, 57.58; H, 6.78.

X-ray Crystallographic Analysis of 13b (with L. J. Todaro and A.-M. Chiu). Crystals of 13b, C₁₃H₁₈O₄S, are orthorhombic, space group P2₁2₁2₁, with *a* = 7.757 (2) Å, *b* = 10.734 (5) Å, *c* = 16.141 (2) Å, *Z* = 4, *d*_{calcd} = 1.336 g cm⁻³, and μ(Cu Kα) = 21.4 cm⁻¹. The structure was solved by a multiple-solution procedure¹³ and was refined by full-matrix least-squares methods. By Hamilton's test,¹⁴ the configuration shown corresponds to the absolute configuration.

(R)-3-Methyl-2-[(phenylsulfonyl)methyl]-1,3-butanediol (14) from 13b. Bromodimethylborane (2.5 mL, 25.6 mmol) was added dropwise, under Ar, to a solution at -78 °C containing 13b (3.5 g, 13.0 mmol) and *N,N*-diisopropylethylamine (0.26 mL) in dry CH₂Cl₂ (130 mL). The resulting mixture was stirred for 2.5 h and then transferred via a cannula to a stirred mixture of aqueous saturated NaHCO₃ (50 mL) and THF (100 mL). The organic phase was separated, and the aqueous residue was extracted with CH₂Cl₂ (2 × 50 mL). The combined extracts were washed with brine, dried (Na₂SO₄), and evaporated to furnish a 1:1 mixture of product and starting material. Chromatography of the residue over silica gel and elution with 3:2:5 EtOAc–CH₂Cl₂–hexane gave after concentration of the appropriate fractions 1.48 g (44%) of 14 as a colorless gum: [α]_D²⁵ -22.5° (CHCl₃, c = 1.042); IR 3625, 3510, 1145 cm⁻¹; UV (EtOH) λ_{max} 216 (ε = 8300) nm; ¹H NMR δ 1.18, 1.28 (2 s, 3 H each, (CH₃)₂C), 2.17 (1 H, m, CH), 2.38 (1 H, t, *J* = 5 Hz, OH), 2.46 (1 H, s, OH), 3.40 (2 H, m, CH₂SO₂), 4.04 (2 H, m, CH₂O), 7.59 (2 H, t, *m*-ArH), 7.68 (1 H, t, *p*-ArH), 7.95 (2 H, d, *o*-ArH); MS *m/z* 259 (32, M⁺ + H). Anal. Calcd for C₁₂H₁₈O₄S: C, 55.79; H, 7.02. Found: C, 55.46; H, 7.01.

(S)-3,3-Dimethyloxiranemethanol (17). A mixture of 16 (5.0 g, 19.9 mmol) and a premixed solution containing MeOH (20 mL) and aqueous 2 N NaOH (20 mL) was stirred at rt for 15 min. Concentration of the mixture gave a slurry which was taken up in CH₂Cl₂ (200 mL), dried (MgSO₄), filtered, and evaporated to provide 3.1 g of product as a colorless thick oil. Purification by column chromatography over silica gel with 30%–50% EtOAc–hexane as eluent furnished 1.36 g (40%) of the known epoxide: [α]_D²⁵ = -21.8° (CHCl₃, c = 1.018) (lit.^{9b} [α]_D²⁵ = -20.1° (CHCl₃, c = 0.42)).

(S)-3-Methyl-2-[(phenylsulfonyl)methyl]-1,3-butanediol (14) from 17. To a stirred solution of CH₂SO₂Ph (2.2 g, 14 mmol) and 22 (1.44 g, 14 mmol) in dry THF (80 mL) at 0 °C was added slowly *n*-BuLi (18.5 mL, 2.5 N in hexane, 18.5 mmol, 46 mmol) to give a yellow suspension. After the addition was complete, the cold bath was removed and the suspension was allowed to stir at rt overnight to give a pale brown homogeneous solution. The reaction mixture then was quenched with a few drops of H₂O and concentrated. The residue was partitioned between EtOAc and H₂O, dried (MgSO₄), filtered, and evaporated. The crude product was chromatographed over silica gel using 20%–30%–50% ethyl acetate–hexane as eluent to give 1.8 g (50% yield) of sulfone diol as a gum [α]_D²⁵ -20.96° (CHCl₃, c = 0.825). All spectroscopic data were identical to the above sample prepared by the enzymatic method. Anal. Calcd for C₁₂H₁₈O₄S: C, 55.79; H, 7.02; S, 12.41. Found: C, 55.56; H, 7.14, S, 12.49.

(S)-2,2,4,4-Tetramethyl-5-[(phenylsulfonyl)methyl]-1,3-dioxane (15). A solution of 14 (1.8 g, 7.0 mmol) in C₆H₆ (50 mL) was mixed with a 10-fold excess of 2,2-dimethoxypropane (7.3 g). A catalytic amount of *p*-TsOH (20 mg) was added, and the reaction mixture was stirred overnight at rt under Ar. The reaction mixture was taken up in EtOAc (50 mL), washed with saturated NaHCO₃ and brine, dried (Na₂SO₄), filtered, and evaporated. The crude product was chromatographed over silica gel and eluted

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(14) Hamilton, W. C. *Acta Crystallogr.* 1965, 18, 502.

with 10%–20%–30% EtOAc in hexane to give, after evaporation of appropriate fractions, 1.5 g (72%) of **15** as a crystalline solid: mp 87–89 °C; $[\alpha]_D^{25}$ –31.46 (CHCl₃, $c = 1.0234$); IR (CHCl₃) 3625, 3510, 1382, 1372, 1138 cm⁻¹; UV (EtOH) λ_{\max} 217 nm ($\epsilon = 8740$); NMR δ 1.15 (3 H, s), 1.18 (3 H, s), 1.38 (6 H, s), 2.17 (1 H, m, –CH(–), 3.06 (2 H, m, –CH₂SO₂), 3.79 (1 H, dd, $J = 8, 12$ Hz, –CH_AO–), 4.15 (1 H, dd, $J = 4.5, 12$ Hz, –CH_BO–), 7.60 (2 H, t, $J = 7.5$ Hz, m -ArH), 7.69 (1 H, t, $J = 7.5$ Hz, p -ArH), 7.95 (2 H, d, $J = 7.5$ Hz, o -ArH). Anal. Calcd for C₁₅H₂₂O₄S: C, 60.38; H, 7.43; S, 10.74. Found: C, 60.20; H, 7.54; S, 10.63.

(1 α ,3 β ,5E,7E)-1,3-Bis[(1,1-Dimethylethyl)dimethylsilyl]oxy]-25,28-[(1-methylethylidene)bis(oxy)]-23-phenylsulfonol-9,10-secocholesta-5,7,10(19)-trien-22-ol (23). A solution of freshly prepared LDA (from 5.04 mL of a 2.5 M solution of *n*-BuLi in hexane and 1.75 mL of *i*-Pr₂NH in dry THF (20 mL) was added to solution of **15** (2.66 g, 8.91 mmol) in dry THF (20 mL) at –70 °C. The resulting mixture was stirred at –70 °C for 30 min and turned yellow when a solution of **7** (5.94 g, 10.37 mmol) in dry THF (20 mL) was added dropwise. Stirring was continued at –70 °C for an additional 3 h after which 2.0 mL of saturated NH₄Cl solution (20 mL) was added dropwise, and the cooling bath was removed. The mixture was poured into saturated NH₄Cl solution (150 mL) and was extracted with EtOAc (2 × 250 mL). The extracts were washed with H₂O, dried (MgSO₄), and evaporated. Chromatography of the residue over 155 g of silica gel packed in hexane and elution with 1% EtOAc in hexane, followed by 10% EtOAc in hexane, gave after concentration of the appropriate fractions 3.10 g (39.9%) of **23** as a gum; IR (CHCl₃) 3547, 1145 cm⁻¹; UV (EtOH) λ 210 ($\epsilon = 21590$), 270 ($\epsilon = 26400$) nm; ¹H NMR (CDCl₃) δ 0.065 (12 H, s, 2 × Me₂Si), 0.56 (3 H, s, CH₃-18), 0.86 (9 H, s, *t*-BuSi), 0.88 (3 H, s, CH₃), 0.89 (9 H, s, *t*-BuSi), 1.20 (3 H, d, $J = 6.5$ Hz, CH₃-21), 1.26 (3 H, s, CH₃-26), 1.34 (3 H, s, CH₃-27), 1.39 (3 H, s), 3.29 (1 H, s), 3.43 (1 H, d, $J = 9$ Hz), 4.05 (1 H, dd, $J = 6, 13$ Hz, CH_A-28), 4.12 (1 H, dd, $J = 6, 13$ Hz, CH_B-28), 4.20 (1 H, br s, CH-3), 4.35 (1 H, br d, $J = 7$ Hz, CH-23), 4.54 (1 H, br s, CH-1), 4.93 (1 H, s, CH_A-19), 4.99 (1 H, s, CH_B-19), 5.82 (1 H, d, $J = 11$ Hz, CH-7), 6.45 (1 H, d, $J = 11$ Hz, CH-6), 7.59 (2 H, t, $J = 7$ Hz, m -ArH), 7.65 (1 H, t, $J = 7$ Hz, p -ArH), 7.96 (2 H, d, $J = 7$ Hz, o -ArH); MS (FAB) 870 (M⁺). Anal. Calcd for C₄₉H₈₂O₇SSi₂: C, 67.54; H, 9.48; S, 3.68; Si, 6.45. Found: C, 67.54; H, 9.64; S, 3.44; Si, 6.31.

[[1 α ,3 β ,5E,7E,22E)-25,28-[(1-Methylethylidene)bis(oxy)]-9,10-secoergosta-5,7,10(19),22-tetraene-1,3-diyl]bis(oxy)]bis-[(1,1-dimethylethyl)dimethylsilyl]silane (24a). A solution of hydroxysulfone **23** (4.09 g, 4.69 mmol) in MeOH (350 mL) was treated with Na₂HPO₄ (15.43 g) and 5% Na–Hg (13.82 g). The mixture was stirred at room temperature for 6 h and filtered through Celite and the filtrate evaporated. The residue was partitioned between 150 mL of EtOAc (150 mL) and H₂O (75 mL). The organic phase was separated, dried (MgSO₄), and evaporated to give 1.92 g of a gum, which was chromatographed over 38 g of silica gel (0–15% EtOAc in hexanes) to give 850 mg after concentration of appropriate fractions (25.4% yield) of **24a**: IR (CHCl₃) 1386, 836 cm⁻¹; UV (EtOH) λ_{\max} 269 ($\epsilon = 27\ 000$) nm; ¹H NMR (CDCl₃) δ 0.05 (6 H, s, Me₂Si), 0.06 (6 H, s, Me₂Si), 0.58 (3 H, s, CH₃-18), 0.86 (9 H, s, *t*-BuSi), 1.02 (3 H, d, $J = 7$ Hz, CH₃-21), 1.20 (3 H, s, CH₃-26), 1.23 (3 H, s, CH₃-27), 1.39 (3 H, s), 1.45 (3 H, s), 3.68 (1 H, dd, $J = 7, 12$ Hz, CH_A-28), 3.80 (1 H, dd, $J = 7, 12$ Hz, CH_B-28), 4.21 (1 H, s, CH-3), 4.53 (1 H, d, $J = 5$ Hz, CH-1), 4.94 (1 H, s, CH_A-19), 4.98 (1 H, s, CH_B-19), 5.20 (1 H, dd, $J = 6, 15$ Hz, CH-23), 5.40 (1 H, dd, $J = 6, 15$ Hz, CH-22), 5.81 (1 H, d, $J = 12$ Hz, CH-7), 6.45 (1 H, d, $J = 12$ Hz, CH-6); MS (FAB) m/z 712. Anal. Calcd for C₄₃H₇₆O₄Si₂: C, 72.41; H, 10.74; Si, 7.88. Found: C, 72.24; H, 10.55; Si, 7.79.

(1 α ,3 β ,5E,7E,22E)-25,28-[(1-Methylethylidene)bis(oxy)]-9,10-secoergosta-5,7,10(19),22-tetraene-1,3-diol (24b). A stirred solution of **24a** (830 mg, 1.16 mmol) in THF (30 mL) was treated,

under Ar, with 28.0 mL of 1.0 M Bu₄F⁺ in THF (28 mL). The mixture was stirred at rt for 5 h, the solvent was evaporated, and the residue was extracted with EtOAc (100 mL) and brine (50 mL). Extractive workup with EtOAc gave a solid, which was chromatographed (0–50% EtOAc in hexanes) over 30 g of silica gel to give 520 mg (92.2%) of **24b**: mp 127–130 °C; $[\alpha]_D^{25}$ +108.6° (EtOAc, $c = 0.638$); IR (CHCl₃) 3607 cm⁻¹; UV (EtOH) λ_{\max} 272 ($\epsilon = 25\ 200$) nm; ¹H NMR (CDCl₃) δ 0.56 (3 H, s, CH₃-18), 1.02 (3 H, d, $J = 7$ Hz, CH₃-21), 1.20 (3 H, s, CH₃-26), 1.25 (3 H, s, CH₃-27), 1.39 (3 H, s), 1.45 (3 H, s), 3.70 (1 H, dd, $J = 6, 11$, CH_A-28), 3.80 (1 H, dd, CH_B-28), 4.32 (1 H, br s, CH-3), 4.50 (1 H, br s, CH-1), 4.98 (1 H, s, CH_A-19), 5.08 (1 H, s, CH_B-19), 5.13 (1 H, dd, $J = 6, 16$ Hz, CH-22), 5.40 (1 H, dd, $J = 6, 16$, CH-23), 5.89 (1 H, d, $J = 12$, CH-7), 6.56 (1 H, d, $J = 12$ Hz, CH-6); MS m/z 484 (2, M⁺). Anal. Calcd for C₃₁H₄₈O₄: C, 76.75; H, 10.00. Found: C, 76.47; H, 10.28.

(1 α ,3 β ,5E,7E,22E)-9,10-Secoergosta-5,7,10(19),22-tetraene-1,3,25,28-tetrol (25). A solution of **24b** (420 mg, 0.866 mmol) in CH₂Cl₂ (28 mL) and MeOH (290 mL) was stirred under Ar with Dowex 50W-X4 ion-exchange resin (11.6 g, H⁺ form, prewashed with methanol) for 15 min. Et₃N (2.0 mL) was added, and the mixture was filtered. Evaporation of the filtrate and chromatography of the residue over silica gel (0–100% EtOAc in hexanes) gave 240 mg (62.3%) of **25** as an amorphous solid: $[\alpha]_D^{25}$ +175.9° (CH₃OH, $c = 0.1137$); IR (CHCl₃) 3606 cm⁻¹; UV (EtOH) λ_{\max} 272 ($\epsilon = 14\ 140$) nm; ¹H NMR (CDCl₃) δ 0.58 (3 H, s, CH₃-18), 1.04 (3 H, d, $J = 6$ Hz, CH₃-21), 1.25 (6 H, s, CH₃-25 + CH₃-26), 2.85 (1 H, s, OH), 2.90 (1 H, s, OH), 3.65 (1 H, dd, $J = 4, 11$ Hz, CH_A-28), 3.85 (1 H, dd, $J = 4, 11$ Hz, CH_B-28), 4.25 (1 H, s, CH-3), 4.50 (1 H, s, CH-1), 4.98 (1 H, s, CH_A-19), 5.13 (1 H, s, CH_B-19), 5.25 (1 H, dd, $J = 7, 14$ Hz, CH-22), 5.44 (1 H, dd, $J = 7, 14$ Hz, CH-22), 5.85 (1 H, d, $J = 12$ Hz, CH-7), 6.60 (1 H, d, $J =$ CH-6);

(1 α ,3 β ,5Z,7E,22E)-9,10-Secoergosta-5,7,10(19),22-tetraene-1,3,25,28-tetrol or 1 α ,25,28-(OH)₃ Vitamin-D₂ (1). A mixture of **25** (33 mg, 0.074 mmol) and 9-acetylanthracene (1.5 mg) in methanol (1 L) was cooled to 0 °C. Ar was passed through the solution, and irradiation was carried out with a 450-W medium-pressure lamp through a uranium filter for 15 min. Evaporation and chromatography of the residue over silica gel gave 28 mg (85%) of **1** as an amorphous solid: $[\alpha]_D^{25}$ +58.9° (CH₃OH, $c = 0.537$) (lit.² $[\alpha]_D^{25}$ +53.5 (EtOH, $c = 0.5$)); IR (CHCl₃) 3608 cm⁻¹; UV (EtOH) λ_{\max} 263 ($\epsilon = 13\ 540$) nm; ¹H NMR (CDCl₃) δ 0.56 (3 H, s, CH₃-18), 1.04 (3 H, d, $J = 6$ Hz, CH₃-21), 1.25 (6 H, s, CH₃-26 + CH₃-27), 2.55 (1 H, t, OH), 3.65 (1 H, dd, $J = 6, 13$ Hz, CH_A-28), 3.74 (1 H, s, OH), 3.83 (1 H, dd, $J = 6, 13$ Hz, CH_B-28), 4.22 (1 H, br s, CH-3), 4.45 (1 H, br s, CH-1), 4.99 (1 H, s, CH_A-19), 5.20 (1 H, dd, $J = 6, 14$ Hz, CH-23), 5.32 (1 H, s, CH_B-19), 5.45 (1 H, dd, $J = 6, 14$ Hz, CH-22), 6.02 (1 H, d, $J = 12$ Hz, CH-7), 6.36 (1 H, d, $J = 12$ Hz, CH-6); MS m/z 445 (M⁺ + H). An analytical sample was prepared by recrystallization from MeO-CHO to give a white solid: mp 171–173 °C (lit.² mp 173–174 °C). Anal. Calcd for C₂₈H₄₄O₄: C, 75.63; H, 9.97. Found: C, 75.46; H, 9.70.

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Supplementary Material Available: The details of crystallography, X-ray analysis, and tables of crystallographic data for **13b** (3 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.